**Red Kite Veterinary Consultants Ltd**

**Home Office Accredited Training**

**Notes for candidates**

1. **Learning outcomes**
2. **Course materials**
3. **Practical sessions**
   1. **Minor procedures**
      1. **Learning Outcomes**
      2. **Required tasks**
      3. **Competence checklist**
   2. **Handling**
      1. **Learning Outcomes**
      2. **Required tasks**
      3. **Assessment**
4. **Written examination**
   1. **Format of the exam**
   2. **Policy on cheating and plagiarism**
   3. **Example questions**
5. **Appendices**
6. **Learning outcomes**

Learning outcomes for accredited training are described in the National Competent Authorities for the implementation of Directive 2010/63/EU on the protection of animals used for scientific purposes working document on the development of a common education and training framework to fulfil the requirements under the Directive. This divides training for personal licence applicants into 11 modules, with further modules being required for project licence applicants. The Home Office has grouped these EU modules into 5 modules for personal licence applicants, as follows:

**Home Office module EU module**

E1L 1 – National legislation

2 – Ethics level 1

PILA 3 – Basic and appropriate biology

4 – Animal care, health and management

5 – Recognition of pain suffering and distress

7 – Minimally invasive procedures without anaesthesia (theory)

8 - Minimally invasive procedures without anaesthesia (skills)

PILK 6 – Humane methods of killing

PILB 20 – Anaesthesia for minor procedures

PILC 21 – Recovery anaesthesia

22 – Surgical procedures

1. **Course materials**

Course materials are available for registered candidates to download from our website. These may be used for personal study but may not be distributed without permission from Red Kite.

Materials may be accessed by visiting [www.redkitevets.co.uk/training](http://www.redkitevets.co.uk/training)

Scroll to online course materials. Select the appropriate course. A password will be required to access the materials.

1. **Practical sessions**

There are a number of practical sessions during these courses, during which you will be assessed. In this section you will find information on the content and format of each practical session and what will be expected of you. It is strongly recommended that you gain some practical handling experience prior to attending the course.

* 1. **Minor procedures (EU module 8)**
     1. Learning Outcomes

The learning outcomes for this module are:

*8.1. Select and explain the best methods for common procedures (such as blood sampling and application of substances) including route/volume/ frequency as appropriate.*

*8.2. Demonstrate that s/he can handle and restrain the animal in the best position for the technique.*

*8.3. Perform minor techniques under supervision, in a manner that does not inflict unnecessary pain, suffering, distress or lasting harm.*

* + 1. Required tasks

The tutor will discuss of health and safety issues surrounding use of sharps and how to maintain sterility. Different needle sizes and suitability for different species will also be discussed. The tutor will demonstrate safe handling of needles and syringes and how to fill syringes, and then demonstrate how to hold an animal (mouse cadaver) for different injections. The tutor will demonstrate i.p., s.c. and i.v. routes.

Task: You will prepare syringes and needles for injections, before demonstrating appropriate handling using cadavers and perform as a minimum ip and sc injections. After injections, you will dispose of the sharps safely.

Additional task (optional): Perform iv injection on cadaver or simulator.

Additional task large animals (optional): Identify injection sites in larger animals and perform ‘mock’ injection (no needle). The tutor will then describe and demonstrate how to carry out cervical dislocation. You will be given the opportunity to perform this procedure on cadavers.

* + 1. Competence checklist

The task will be assessed according to the competence checklist at appendix 2. A second tutor will normally be present during the practical, in which case this tutor may perform the assessment.

* 1. **Handling**
     1. Learning Outcomes

The learning outcome for this module is: *Be able to approach, handle/pick up and restrain an animal and return it to its cage/pen in a calm, confident and empathetic manner such that the animal is not distressed or caused harm.*

* + 1. Required tasks

Handling will be demonstrated by the trainer and you will then be asked to perform the following task. You will be given a number of opportunities to carry out the task to the required standard subject to your skill level and the animal stress level. You may be asked some additional questions to demonstrate that you understand why you are carrying out the task in a particular way. A second trainer will usually be present during the training session and in this case the second trainer will perform the assessment.

Task: small animals/poultry. Candidate removes cage from rack if appropriate. Candidate selects appropriate animal and removes from cage/pen. Candidate transfers animal from one cage to another using an appropriate method (optional – depends on species). Candidate restrains animal in a manner appropriate for examination or minor procedure. Candidate returns animal to cage/pen and replaces cage on rack in an appropriate manner.

Task: large animals. Candidate enters pen and approaches animal. Candidate restrains animal in a manner appropriate for examination or minor procedure. Candidate releases animal and leaves pen.

* + 1. Assessment

The task will be assessed according to the competence checklist at appendix 3, and marked using the assessment grid. You need only achieve a grade of ‘pass’. If you do not achieve this standard in any one category during the training session you will be expected to undertake further practice in your home institution before retaking the assessment.

1. **Written examination**
   1. **Format of the exam – personal licence applicant category A and B.**

The examination consists of different papers for each core module (E1L, PILA, PILB, PILK).

For **E1L**, there is a preliminary paper consisting of 5 multiple choice questions on the legislation and responsibilities of the personal licence holder. **You must score 100% on this section**. If you do not score 100% the rest of the paper will not be marked and you will need to resit this part of the exam.

All other modules consist of questions similar to the example questions shown below. You must score 70% in these papers to pass.

You will have 2 minutes per question to complete the assessment. Reference to course materials or use of the internet is forbidden during the written examination.

Candidates that fail to reach the desired standard will be permitted to retake an assessment on a single occasion. If they fail twice they may be required to resit the course.

* 1. **Policy on cheating and plagiarism**

It is unacceptable for a candidate to infringe, or attempt to infringe, the regulations governing the conduct of examinations or to engage, or attempt to engage, in conduct for the purpose of gaining an unfair advantage with a view to obtaining a better result than he or she would otherwise achieve.

Examples of these are:

1. Copying from the examination script of another candidate

2. Impersonation of others

3. Fabrication of results

4. Plagiarism

5. Collusion

6. Contract cheating

7. Having inadmissible material on your person during the examination, e.g., mobile phone, other electronic device, course notes.

Candidates believed to be cheating may be asked to leave the examination and their papers will not be marked. They may then have to wait for a period of three months before repeating the examination.

* 1. **Example questions**

|  |  |
| --- | --- |
| **Question : 5 marks**  You have experimental animals on a study which is due to continue for several weeks; during this time, you plan to go on holiday for a week. What should you do before you go? | |
| Nothing, the NACWO will monitor them in your absence |  |
| Kill them all before you leave and terminate the study |  |
| Notify the Establishment licence holder so they can sort out alternative cover |  |
| Arrange with another personal licensee to take responsibility in your absence | x |
| Ask an experienced member of the animal care team to check on them every few days and keep a record |  |

* + 1. **Example L killer question**
    2. **Example L general question**

|  |  |
| --- | --- |
| **Question : 5 marks**  Identify with a T(rue) or F(alse) the duties of an Establishment Licence Holder as required by the ASPA. | |
| To provide practical assistance to licence applicants | F |
| To authorise the use of controlled drugs | F |
| To ensure the establishment is appropriately staffed | T |
| To comply with the Health & Safety at Work Act | F |
| To countersign project licence applications | T |

* + 1. **Example PILA question**

|  |  |
| --- | --- |
| **Question : 5 marks**  The severity of procedures is determined by the degree of pain, suffering, distress or lasting harm expected to be experienced by an individual animal during the course of the procedure. The final severity classification takes account of additional factors, assessed on a case by case basis. Identify as T(rue) or F(alse) the following statements: | |
| The training experience of an animal with respect to the procedure is taken into account | T |
| The type of species or its genotype are not taken into account | F |
| Procedures performed entirely under terminal general anaesthesia are termed non-recovery | T |
| Cumulative suffering must be factored into to the final assessment | T |
| The maturity, age and gender of the animal is irrelevant | F |

**Appendix 1: Learning outcomes for accredited training**

1. **E1L:** 
   1. **Module 1: National legislation**

Trainees should be able to:

1.1. Identify and describe the national and EU laws and guidance which regulate the scientific use of animals and in particular the activities of those carrying out scientific procedures involving them.

1.2. Identify and describe related animal welfare legislation.

1.3 Describe the authorisation that is needed before acting as user, breeder or supplier of laboratory animals and especially the authorisation required for projects and where applicable individuals.

1.4. List sources of information and support that are available (regarding national legislation).

1.5. Describe the role of the personnel mentioned in Article 24, 25 and 26, and their statutory duties and other responsibilities under the National Legislation.

1.6. Describe the roles and responsibilities of the local animal welfare bodies and the national committee for the protection of animals used for scientific purposes.

1.7. Indicate who is responsible for compliance at an establishment and how this responsibility may be exercised (e.g. through the local AWB).

1.8. Describe when a procedure becomes regulated under National legislation (minimum threshold of pain, suffering, distress or lasting harm).

1.9. Indicate who bears primary responsibility for the animals undergoing procedures.

1.10. List which species, including respective stages of development that are included in the scope of the Directive / National law.

1.11. Indicate the circumstances in which animals under the scope of the Directive should be humanely killed or removed from the study to receive veterinary treatment.

1.12. Describe the legislative controls over the killing of animals bred or used for scientific procedures

**b. Module 2: Ethics, animal welfare and the Three Rs (level 1)**

Trainees should be able to:

2.1. Describe the differing views, within society, relating to the scientific uses of animals and recognise the need to respect these.

2.2. Describe the responsibility of humans when working with research animals and recognise the importance of having a respectful and humane attitude towards working with animals in research.

2.3. Identify ethical and animal welfare issues in their own work and be aware and able to reflect on the consequences of their own actions.

2.4. Recognise that compliance with ethical principles may contribute to the long-term trust and acceptance in scientific research from the general public.

2.5. Describe how the law is based on an ethical framework which requires 1) weighing the harms and benefits of projects (the harm/benefit assessment) 2) applying the Three Rs to minimise the harm, maximise benefits and 3) promote good animal welfare practices.

2.6. Describe and discuss the importance of the ThreeRs as a guiding principle in the use of animals in scientific procedures.

2.7. Explain the Five Freedoms and how these apply to laboratory species

2.8. Describe the concept of harms to animals including avoidable and unavoidable suffering, direct, contingent and cumulative suffering

2.9. Describe the severity classification system, and give examples of each category. Describe cumulative severity and the effect this may have on the severity classification.

2.10. Describe the regulations regarding re-use of animals.

2.11. Describe the importance of good animal welfare including its effect on scientific outcomes as well as for societal and moral reasons.

2.12. Describe the need for a culture of care and the individual’s role in contributing to this.

2.13. Describe relevant sources of information relating to ethics, animal welfare and the implementation of the Three Rs.

2.14. Be aware of different search tools (e.g. EURL ECVAM Search Guide, Go3Rs) and methods of search (e.g. Systematic reviews, meta analysis).

1. **PILA**
   1. **Module 3.1: Basic and appropriate biology - theory**

Trainees should be able to:

3.1.1. Describe basic anatomy, physiology, reproduction and behaviour of the relevant species.

3.1.2. Recognize and describe life events that have the potential to cause suffering including sourcing, transport, housing, husbandry, handling and procedures (on a basic level).

3.1.3. Indicate how good welfare can promote good science: e.g. explain how the failure to attend to biological and behavioural needs may affect the outcome of procedures.

3.1.4. Indicate how husbandry and care may influence experimental outcome and the number of animals needed e.g. example where the place in the room influences the outcome, hence randomisation.

3.1.5. Describe the dietary requirements of the relevant animal species and explain how these can be met.

3.1.6. Describe the importance of providing an enriched environment (appropriate to both the species and the science) including social housing and opportunities for exercise, resting and sleeping.

3.1.7. When relevant to the species, recognise that there are different strains, and that these can have different characteristics which can affect both welfare and science.

3.1.8. When relevant to the species, recognise that alterations to the genome can affect the phenotype in unexpected and subtle ways, and the importance of monitoring such animals very carefully.

3.1.9. Maintain and interpret accurate, comprehensive records of animals held in the animal facility, including the wellbeing of the animals

**b. Module 3.2: Basic and appropriate biology – skills**

3.2.1. Be able to approach, handle/pick up and restrain an animal and return it to its cage/pen in a calm, confident and empathetic manner such that the animal is not distressed or caused harm.

**c. Module 4: Animal care, health and management – species specific (theory)**

Trainees should be able to:

4.1. Describe suitable routines and husbandry practices for the maintenance, care and welfare for a range of animals used in research, to include small laboratory species and large animal species where appropriate.

4.2. Describe suitable environmental and housing conditions for laboratory animals, how conditions are monitored and identify the consequences for the animal resulting from inappropriate environmental conditions.

4.3. Recognise that changes to or disruption of circadian or photoperiod can effect animals.

4.4. Describe the biological consequences of acclimatisation, habituation and training

4.5. Describe how the animal facility is organized to maintain an appropriate health status for the animals and the scientific procedures.

4.6. Describe how to provide water and an appropriate diet for laboratory animals including the sourcing, storage and presentation of suitable foodstuffs and water

4.7. List the methods, and demonstrate an understanding of appropriate, safe and humane handling, sexing and restraint of one or more named species for common scientific procedures.

4.8. Name different methods for marking individual animals and state an advantages and disadvantage for each method.

4.9. List potential disease risks in the animal facility, including specific predisposing factors which may be relevant. Name methods available for maintaining appropriate health status (including use of barriers, different containment levels use of sentinels as relevant to the species).

4.10. Describe appropriate breeding programmes

4.11. Describe how genetically altered animals can be used for scientific research and the importance of monitoring such animals very carefully.

4.12. List the correct procedures for ensuring health, welfare and care of animals during their transport.

4.13. List potential human health hazards associated with contact with laboratory animals (including allergy, injury, infection, zoonosis) and how these can be prevented.

**d. Module 5: Recognition of pain, suffering and distress**

Trainees should be able to:

5.1. Recognise normal or desirable behaviour and appearance of the individuals in the context of species, environment and physiological status.

5.2. Recognise abnormal behaviour and signs of discomfort, pain, suffering, or distress, as well as signs of positive well-being and principles of how pain, suffering and distress can be managed.

5.3. Discuss factors to be considered and methods available for assessing and recording the welfare of animals e.g. score sheets.

5.4. Describe what a humane end point is. Identify criteria to be used to set humane endpoints. Define action to be taken when a humane endpoint is reached and consider possible options for refining methods to finish at an earlier endpoint.

5.5. Describe the severity classifications included in the Directive and give examples of each category; explain cumulative severity and the effect this may have on the severity classification.

5.6. Describe the circumstances when anaesthesia or analgesia may be necessary to minimise pain, suffering, distress or lasting harm

1. **Module 7: Minimally invasive procedures without anaesthesia – (theory)**

Trainees should be able to:

7.1. Describe appropriate methods and principles to be followed when handling animals (including methods of manual restraint and use of restricted environments).

7.2. Describe the biological impact of procedures and restraint on physiology.

7.3. Describe refinement opportunities for procedures and restraint e.g. through training (using positive re-enforcement), habituation and socialisation of animals.

7.4. Describe techniques/procedures including, for example, injection, sampling and dosing techniques (routes/volumes/frequency), dietary modification, gavage, tissue biopsy, behavioural tests, use of metabolic cages.

7.5. Describe how to perform minor techniques and relate appropriate sample volumes and sampling frequencies for the relevant species.

7.6. Describe the need for rigour and consistency in conducting scientific procedures and the correct recording and handling of samples.

7.7. Describe appropriate methods for the assessment of the welfare of animals with respect to the severity of procedures and know what appropriate action to take.

7.8. Recognize that refinement is an on-going process and know where to find relevant, up-to-date, information.

7.9. Describe the biological consequences of transport, acclimatization, husbandry conditions and experimental procedures on the species concerned and describe how these can be minimised.

1. **Module 8: Minimally invasive procedures without anaesthesia – skills**

Trainees should be able to:

8.1. Select and explain the best methods for common procedures (such as blood sampling and application of substances) including route/volume/ frequency as appropriate.

8.2. Demonstrate that s/he can handle and restrain the animal in the best position for the technique.

8.3. Perform minor techniques under supervision, in a manner that does not inflict unnecessary pain, suffering, distress or lasting harm.

1. **PILK**
   1. **Module 6.1: Humane methods of killing - theory**

Trainees should be able to:

6.1.1. Describe the principles of humane killing (e.g. what constitutes ‘a good death’)

6.1.2. Describe the different methods by which the relevant animals are allowed to be killed, the influence different methods can have on scientific outcomes, and how to select the most appropriate method.

6.1.3. Explain why someone competent to kill animals should be available at all times (whether care staff or person carrying out procedures)

* 1. **Module 6.2: Humane methods of killing (skills)**

Trainees should be able to:

6.2.1. Proficiently and humanely carry out euthanasia using appropriate techniques on relevant species of laboratory animals

6.2.2. Demonstrate how death is confirmed and how cadavers should be processed or otherwise disposed of.

1. **PILB**
   1. **Module 20: Anaesthesia for minor procedures**

Trainees should be able to:

20.1. Define sedation, local and general anaesthesia

20.2. Identify the three components of the triad of anaesthesia and understand that different anaesthetic agents produce these to different degrees.

20.3. Define balanced anaesthesia and indicate that this is best achieved by using drugs in combinations to achieve all components of the anaesthetic triad to an acceptable degree

20.4. Relate why and when sedation or anaesthesia might be used for restraint.

20.5. List the factors to be considered in pre-anaesthetic evaluation of animals - how to perform a basic health check, consider physiological or pathological status of the model they are working with and how these may influence the choice of anaesthetic agent.

20.6. Discuss the relative merits / drawbacks and principles of selection of different agents and their application, including calculation of doses, in relevant species, including injectable and volatile agents (or dissolved agents in the case of aquatic species), including local anaesthesia regimes

20.7. Indicate the importance of minimising stress prior to anaesthesia in reducing the likelihood of complications due to anaesthesia.

20.8. Recognise when premedication is beneficial to incorporate into an anaesthetic regime.

20.9. Describe and demonstrate the correct set-up, operation and maintenance of anaesthetic equipment appropriate to the species concerned.

20.10. Evaluate and appreciate the different levels and planes of anaesthesia (voluntary excitement, involuntary excitement, surgical anaesthesia (light, medium & deep), excessively deep).

20.11. List the factors indicating that an animal is suitably anaesthetized (stable and of appropriate depth) to enable procedures to be undertaken and what actions should be taken if an adverse event occurs. This will include basic “hands on” and “observational” anaesthetic monitoring techniques, including assessment of reflexes appropriate for species.

20.12. Describe methods of optimising post anaesthetic recovery (e.g. heat blankets, analgesia, reversal agents, access to food and water, environmental conditions) to ensure a smooth and rapid recovery from anaesthesia.

20.13. Demonstrate an understanding of safe / good working practices with regard to use, storage and disposal of anaesthetic and analgesic agents.

1. **PILC**
   1. **Module 21: Advanced anaesthesia for surgical or prolonged procedures**

Trainees should be able to:

21.1. Relate why and when anaesthesia might be used, including additional factors relevant for long term anaesthesia.

21.2. Relate the need for and list the factors to be considered in pre-anaesthetic evaluation of animals, including acclimatisation.

21.3. Discuss the use of pre-anaesthetic agents and analgesics as part of a balanced anaesthetic regime.

21.4. Indicate that a range of drugs are commonly used for premedication and the induction and maintenance of anaesthesia in relevant laboratory species, and identify where to get advice on the different drug available and their use.

21.5. Describe how an animal’s concurrent pathology may require specific anaesthetic regimen, monitoring or nursing care.

21.6. Indicate types of agents used for the induction and maintenance of general anaesthesia, their advantages and disadvantages and when each might be used.

21.7. Describe how anaesthetic agents interact to produce the three components of the anaesthetic triad to different degrees, and how balanced anaesthesia might be best achieved by using combinations.

21.8. Demonstrate a sufficient understanding of anaesthetic agents having a low analgesic effect, potentially requesting the use of an additional analgesia.

21.9. List the factors to be considered when monitoring anaesthesia both for anaesthetic depth and physiological stability. Indicate how to determine that an animal is sufficiently deeply anaesthetised to enable painful procedures to be undertaken, and what action should be taken if an adverse event occurs.

21.10. List methods which can used to assist monitoring of anaesthesia (e.g. ECG, BP, Urine output, Oxygen saturation, CO2) and how these can be monitored.

21.11. Monitor anaesthetic depth and the animals' vital signs, using both clinical signs, and electronic apparatus if appropriate.

21.12. Describe and demonstrate the correct set-up, operation and maintenance of anaesthetic and monitoring equipment appropriate to the species concerned.

21.13. Demonstrate competence in maintaining and interpreting records of pre- and post- anaesthetic induction and whilst an animal is anaesthetised, as well as in managing the animal care adequately

21.14. Indicate the problems that may occur during anaesthesia, and understand how to avoid these, or manage them if they occur.

21.15. Demonstrate an understanding of mechanical ventilation.

21.16. Describe methods to optimise post anaesthetic recovery to ensure a smooth and rapid recovery from anaesthesia, as in Basic Module but with additional methods required, including analgesia and fluid replacement, for animals having undergone lengthy anaesthesia of surgical procedure.

21.17. Consider the consequences of anaesthesia and the surgical procedures on recovery.

21.18. Appreciate how the choice of anaesthetic agent will determine the rate of recovery and describe how duration and quality of anaesthesia governs the rate of recovery.

21.19. Describe the problems that can arise (in the post-operative period), and indicate how to avoid these, or manage them if they occur.

21.20. Discuss how to integrate a program of pain management into an overall scheme of perioperative care.

21.21. Indicate some of the problems associated with pain recognition and pain management in animals.

21.22. Demonstrate a sufficiently detailed understanding of analgesics to be able to administer safely, including routes of administration and potential adverse effects.

21.23. Demonstrate an understanding of safe / good working practices with regard to use, storage and disposal of anaesthetic and analgesic agents.

* 1. **Module 22: Principles of surgery**

Trainees should be able to:

22.1. Explain the relevance and need for pre-operative assessment and, where appropriate, conditioning.

22.2. Identify sources of reference for good surgical practice

22.3. Describe the process of tissue healing and relate to this to the importance of asepsis and hygienic practices, wound creation, the principles of tissue handling and selection of a suitable surgical approach

22.4. Discuss possible causes of delayed or impaired wound healing or other post-surgical complications and describe ways in which these can be avoided or, if they occur, treated

22.5. Describe in general terms how personnel, animals, instruments and equipment should be prepared for aseptic surgery

22.6. List the principles of successful surgery (e.g. Halstead’s principles) and indicate how to achieve these

22.7. Describe the characteristics of different, commonly-used instruments, suture materials and needles

22.8. Relate the importance of good technique in accessing surgical sites, handling tissues and repairing incisions

22.9. Indicate the characteristics of different suture patterns and their applicability to different situations

22.10. Demonstrate how to place a suture correctly

22.11. Describe common post-surgical complications and their causes

22.12. Relate the principles of post-surgical care and monitoring

22.13. Describe the planning of surgical procedures and discuss the competencies required of all personnel involved

22.14. Demonstrate competence in surgical techniques, including ablations and incisions and their closure by methods appropriate to the tissue concerned

22.15. Describe particular aspects of care appropriate for animals before, during and after surgical or any other potentially painful intervention

**Appendix 2: Competence checklist for module 8, minor procedures**

**Competence checklist: the candidate should**

* Check licence authorities (or state that these need to be checked)
* Select appropriate PPE
* Select appropriate animal
* Handle and restrain animal/cadaver/simulator using appropriate method for the procedure
* Correctly identify injection sites
* Select appropriate equipment:
* Assemble needle and syringe correctly
* Calculate appropriate injection/sample volume
* Prepare animal appropriately
* Either: remove needle and place syringe in correct location for performing procedure
* Or: Correctly perform injection on cadaver or simulator:
* Handle sample appropriately
* Dispose of needle and syringe appropriately
* Apply appropriate method for haemostasis if appropriate
* Return animal/cadaver/simulator to home cage appropriately
* Complete records appropriately

**Appendix 3: Competence checklist and assessment grid for Module 3.2: Handling**

**Competence checklist: the candidate should**

* Check the risk assessment for handling and/or licence authorities if appropriate
* Select appropriate PPE
* Identify correct pen/cage/animal
* **Either** remove cage from rack
  + **or** enter pen\*
* Remove food hopper or water bottle from cage/pen as appropriate
* Approach animal in calm and confident manner
* Assess general state of health of animal
* Select appropriate method of restraint for animal in different situations, e.g. handling for examination, restraining for procedures
* Handle animal safely and securely in appropriate manner as above
* Identify gender of animal
* Release animal safely
* **Either** Replace cage lid and return cage to rack
  + **Or** replace removed items and leave pen ensuring it is secure\*
  + **Or** Candidate replaces food hopper or other items, then candidate leaves pen quietly and calmly and remembers to shut the gate\*
* Candidate checks all animals are still in the pen/cage

**Handling assessment grid**

|  |  |  |  |
| --- | --- | --- | --- |
| Observation | 1.Criteria | 2. Criteria | 3. Criteria |
| Approach | Anxious/tentative  Difficulty in approaching animal | Not confident but approaches animal appropriately | Confident & relaxed  Approaches animal immediately |
| Handling | Unable to demonstrate correct ways to hold animal | Can demonstrate one or more ways to hold but not always securely | Can restrain securely in a number of ways |
| Animal Stress | Animal obviously stressed;  Vocalises and struggles | Animal mostly calm, with some struggling but settles | Animal is calm and relaxed, does not struggle |
| Theory | Candidate demonstrates little or no knowledge | Understands key aspects of handling and restraint | Has a detailed understanding of handling and restraint |
|  | Unsatisfactory | Pass | Good |